



Dietary Phytoncide Supplementation Improved Growth Performance and Meat Quality of Finishing Pigs

Han Lin Li, Pin Yao Zhao, Yan Lei, Md Manik Hossain, Jungsun Kang¹, and In Ho Kim*

Department of Animal Resource and Science, Dankook University, Cheonan 330-714, Korea

ABSTRACT: We conducted this 10-wk experiment to evaluate the effects of dietary phytoncide, Korean pine extract as phytogetic feed additive (PFA), on growth performance, blood characteristics, and meat quality in finishing pigs. A total of 160 pigs ([Landrace×Yorkshire]×Duroc, body weight (BW) = 58.2±1.0 kg) were randomly allocated into 1 of 4 treatments according to their BW and sex, 10 replicate pens per treatment with 4 pigs per pen were used (2 barrows and 2 gilts). Dietary treatments were: CON, control diet; PT2, CON+0.02% PFA; PT4, CON+0.04% PFA; PT6, CON+0.06% PFA. Overall, average daily gain (ADG) was higher in PT4 ($p<0.05$) than in PT6, average daily feed intake (ADFI) was lower in PT6 than in CON ($p<0.05$). Besides ADFI decreased linearly ($p<0.05$) with the increased level of phytoncide and gain:feed ratio in PT4 treatment was higher ($p<0.05$) than CON treatment. During 5 to 10 weeks and overall, quadratic ($p<0.05$) effect was observed in ADG among the treatments. At the end of this experiment, pigs fed with PT4 diet had a greater ($p<0.05$) red blood cell concentration compared to the pigs fed CON diet. Water holding capacity increased linearly ($p<0.05$) with the increased level of phytoncide supplementation. Moreover, firmness, redness, yellowness, and drip loss at day 3 decreased linearly ($p<0.05$) with the increase in the level of phytoncide supplementation. In conclusion, inclusion of phytoncide could enhance growth performance without any adverse effects on meat quality in finishing pigs. (**Key Words:** Blood Profile, Finishing Pig, Growth Performance, Meat Quality, Phytoncide)

INTRODUCTION

Long-term storage of animal feed in high humidity and high temperature condition is a major problem of livestock farm production and management, because nutrients in feed can be decomposed by harmful microorganisms. For example, *Aspergillus* and other fungi may produce toxins such as aflatoxins, while pathogenic bacteria, such as *Salmonella enteritidis* and *Escherichia coli*, present in feed or feedstuffs for animal consumption. All of these can be dangerous for animal health (Cole and Cox, 1981). The diet of pig consists mainly of cereal grains, which often easily contaminated with aflatoxin-producing molds. It is reported that pigs to be one of the most sensitive animals to the effects of aflatoxin (Shi et al., 2005). Thus, aflatoxin-contaminated feed is a serious threat to the swine industry.

* Corresponding Author: In Ho Kim. Tel: +82-41-550-3652, Fax: +82-41-565-2949, E-mail: inhokim@dankook.ac.kr

¹ Genebiotech. Co. Ltd., Seoul 06774, Korea.

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Phytoncide are natural volatile compounds emitted by trees and plants as a protective mechanism against harmful insects, animals, and microorganisms. Trees and plants synthesize phytoncide as secondary metabolites of photosynthesis. Phytoncide are enriched in terpenoids, phenylpropanoids, and alkaloids; major ingredients include monoterpenoids, including α -pinene, careen, and myrcene (Durham et al., 1994; Nose, 2000; Oikawa, 2005). As a mean of suppressing activity of microorganisms, such as bacteria and fungi, relatively large amounts of phytoncide were released by Korean pines and other conifers into the environment (Pečiulytė et al., 2010). The suppressive activity relies on essential oil, terpenoids, alkaloid, and phenylpropanoids of phytoncide (Durham et al., 1994). Due to them, phytogetic feed additive (PFA) have a potential capacity to intercalate into the bacterial membrane, disintegrate the membrane, and cause ion leakage (Burt, 2004; Hashemi and Davoodi, 2010). Moreover, this can interfere with regulatory networks of signal transduction and gene expression in aflatoxin biosynthesis, or by

blocking the enzymatic activity of a biosynthetic enzyme (Holmes et al., 2008).

These previous findings suggested that phytoncide as a potential feed additive. In support of this, one study described the antioxidative potential of phytoncide and their effects in reducing oxidative stress responses in stroke-prone spontaneously hypertensive rats (Kawakami et al., 2004). Moreover, stress level was known to have negative correlation with meat quality (Terlouw, 2005). According to these previous studies, we could hypothesize that dietary phytoncide supplementation may help to improve growth performance and meat quality in finishing pigs. However, the effect of dietary phytoncide supplementation in finishing pigs has not yet been investigated. Thus, this study was conducted to determine the effect of dietary phytoncide supplementation on growing performance, blood characteristics, and meat quality in finishing pigs.

MATERIALS AND METHODS

Preparation of phytoncide and experimental animals

The phytoncide (PHYLUS Company, Seoul, Korea) used in this study was extracted from Korean pine. It contained 20% active substance (flavonoid, phenolic compounds, alkaloid, tannin, terpene, saponin) and 80% carrier (dextrin). About 40% of active substance were essential oils. The experimental protocol was approved by the animal care and use committee of Dankook University (Ethics Approval Number: DK-1152).

A total of 160 finishing pigs ([Landrace×Yorkshire]×Duroc, body weight [BW] = 58.2±1.0 kg) were randomly allocated into 1 of 4 treatments according to their BW and sex. Ten replicate pens per treatment with 4 pigs per pen were used (2 barrows and 2 gilts). Dietary treatments included: CON, control diet; PT2, CON+0.02% PFA; PT4, CON+0.04% PFA; PT6, CON+0.06% PFA. All pigs were housed in pens (1.8×1.8 m) with a self-feeder and nipple drinker that allowed pig *ad libitum* access to feed and water throughout the whole experimental period. Temperature was maintained at 24°C to 26°C. Dietary phytoncide was supplemented in diets by replacing the same amount of corn in our current study. As showed in Table 1, all diets used in present study were formulated to meet or little exceed the nutrient recommendations of NRC (2012). Diets were formulated to the same concentrations of lysine, crude protein (CP), metabolizable energy, calcium, and phosphorus (Table 1).

Sampling and measurements

BW and feed intake were measured at the start of experiment and at the end of 5 and 10 wk to calculate average daily gain (ADG), average daily feed intake (ADFI) and gain:feed. Samples of diets were analyzed for

Table 1. Formula and chemical composition of experimental diet (as-fed basis)

Items	
Ingredients (%)	
Corn	67.81
Soybean meal (48%)	17.88
Rice bran	5.00
Molasses, sugar beets	5.00
Tallow	2.00
Dicalcium phosphate	0.75
Calcium carbonate	0.94
L-Lys ·HCl (78%)	0.30
Salt	0.15
Vit-perimix*	0.07
Min-premix*	0.10
Calculated nutrient composition	
Metabolizable energy (kcal/kg)	3,308
Crude protein (%)	15.63
Lysine (%)	0.97
Calcium (%)	0.60
Phosphorus (%)	0.56
Analyzed nutrient composition	
Metabolizable energy (kcal/kg)	3,301
Crude protein (%)	15.50
Lysine (%)	0.968
Calcium (%)	0.59
Phosphorus (%)	0.52
Crude fat (%)	5.33
Crude fiber (%)	4.14

* Provided per kg of complete diet: 4,000 IU of vitamin A; 800 IU of vitamin D₃; 17 IU of vitamin E; 2 mg of vitamin K; 4 mg of vitamin B₂; 1 mg of vitamin B₆; 16 µg of vitamin B₁₂; 11 mg of pantothenic acid; 20 mg of niacin; 0.02 mg of biotin; 220 mg of Cu (CuSO₄ 5H₂O); 175 mg of Fe (FeSO₄ H₂O); 191 mg of Zn (ZnSO₄ H₂O); 89 mg of Mn (MnO); 0.3 mg of I (CaI₂); 0.5 mg of Co (CoSO₄ 7H₂O); 0.3 mg of Se (Na₂SeO₃ 5H₂O).

CP (procedure 984.13; AOAC, 2006), calcium (procedure 968.08; AOAC, 2006), phosphorus (procedure 946.06; AOAC, 2006), and crude fiber (procedure 978.10; AOAC, 2006). Crude fat were determined according to the method of Thiex et al. (2003) and Kjeldahl N was determined according to the method of Thiex et al. (2002). The concentrations of neutral detergent fiber and acid detergent fiber were determined according to the method of Van Soest et al. (1991). The concentration of crude fiber was analyzed using heat-stable α -amylase and sodium sulfite without correction for insoluble ash as adapted for an Ankom Fiber Analyzer (Ankom Technology, Macedon, NY, USA). Samples of diets were analyzed for metabolizable energy via an adiabatic oxygen bomb calorimeter (Parr Instruments Co., Moline, IL, USA).

Blood samples were collected via jugular vein into either 5-mL vacuum or 5-mL K3EDTA vacuum tube (Becton Dickinson Vacutainer Systems, Franklin Lakes, NJ,

USA) from 2 pigs each pen on the initial and final day of the experimental period. Concentrations of red blood cell (RBC) and white blood cell (WBC) in whole blood samples were measured using the automatic blood analyzer (ADVIA 120, Bayer, Tarrytown, NY, USA). Serum samples were centrifuged (3,000×g) for 15 min at 4°C, and then concentrations of cortisol was measured using the automatic biochemistry analyzer (HITACHI 747, Tokyo, Japan).

At the end of the experiment, 10 pigs per treatment (1 pig per pen with a BW of pen average) were transported to the abattoir for slaughter. The carcasses were placed in a conventional chiller at 4°C. After 24 h chill period, carcasses were fabricated into primal cuts. Meat samples, which included lean and fat, were taken via perpendicular cut loins into 2-cm-thick chop beginning from the 10th and 11th ribs region. Back fat thickness was measuring midline fat thickness by using a real-time ultrasound instrument (Piglot 105, SFK Technology, Herlev, Denmark) according to method of Wang et al. (2009). The pH of longissimus muscle (LM) was measured in 24 H post-mortem with an insertion of glass electrode (Radiometer, Lyon, France) connected to a pH-meter (NWKbinar pH, K-21, Landsberg, Germany). The electrode was calibrated at 20°C in buffers at pH value of 4.00 and 7.00. Surface LM color (Minolta L*,a*,b*) was measured in triplicate on a freshly-cut surface with a Minolta Chromameter (Minolta CR 301, Tokyo, Japan). The water holding capacity (WHC) was measured according to the methods of Kauffman et al. (1986). In brief, 0.2 g sample was pressed at 3,000 psi for 3 min on 125-mm-diameter filter paper. The areas of the pressed sample and expressed moisture were delineated and then determined with a digitizing area-line sensor (MT-10S; M.T. Precision Co. Ltd., 123 Tokyo, Japan). A ratio of water: meat areas was calculated, giving a measure of WHC (the smaller ratio indicate the higher the WHC). The proportion of LM acceptable for Pork Composition and Quality Assessment Procedures (NPPC, 1991) was determined via the selection of LM with acceptable color, firmness, and marbling (all measures 3 or greater, based on a scale of 1 to 5; NPPC, 1991). Drip loss was measured using approximately 2 g of meat sample according to the plastic bag method, which was described by Honikel (1998).

Statistical analyses

For current experiment, effects of treatment (0, 0.02%, 0.04%, 0.06% phytoncide) were analyzed by analysis of variance (ANOVA) using the Mixed Models procedure of SAS (SAS Institute, 2003). The pen was used as the experimental unit for growth performance analysis, while data of blood profile and meat quality were based on individual pigs. The initial BW was used as a covariate for ADFI and ADG, and initial values were used as a covariate

for blood profile. The complete model of blood profiles included the main effects of treatment, litter and time, and the treatment×time interaction. Differences among treatment means were determined using the Tukey's multiple range test. Probabilities less than 0.05 were considered to be significant. After ANOVA analysis, linear and quadratic polynomial contrasts were performed to determine the effects of different level of phytoncide (0.02%, 0.04%, and 0.06%) in the diet.

RESULTS

Growth performance

During 0 to 5 weeks, pigs fed the PT4 diet had a greater ($p<0.05$) ADG than pigs fed the PT6 diet (Table 2). From 5 to 10 weeks, pigs fed the PT4 diet had a greater G:F ($p<0.05$) than pigs fed the CON diet. During the entire experimental period, ADG was increased with supplementation with 0.04% phytoncide ($p<0.05$) compared with the inclusion of 0.06% phytoncide. Furthermore, ADFI in the PT6 group was lower ($p<0.05$) than that in the CON group, and G:F in PT4 treatment was higher ($p<0.05$) than with CON.

Moreover, throughout the experimental period, the ADFI decreased linearly ($p<0.05$) with the increased level of phytoncide, and a significant quadratic effect ($p<0.05$) was observed in G:F among treatments. During 5 to 10 weeks and overall, a significant quadratic effect ($p<0.05$) was observed in ADG among treatments.

Blood profiles

Blood profiles are presented in Table 3. At the end of the experiment, pigs fed the PT4 diet had a higher ($p<0.05$) RBC concentration than the pigs fed the CON diet. Also, the cortisol concentration was decreased ($p<0.05$) with PT2 and PT4 treatment versus the CON treatment. Time effect was observed in cortisol and WBC, while no treatment×time interaction effect was observed. Moreover, a significant quadratic effect was observed in cortisol, RBC, and WBC among treatments.

Meat quality

As shown in Table 4, supplementation with 0.02% and 0.06% phytoncide increased ($p<0.05$) the sensory color compared with CON treatment, and inclusion of 0.04% and 0.06% phytoncide decreased ($p<0.05$) sensory firmness versus CON treatment. WHC in PT6 was higher ($p<0.05$) than that in the PT2 and CON treatments. No difference ($p<0.05$) was observed in loin muscle area, pH, meat color, or drip loss ($p>0.05$). Moreover, firmness, redness, yellowness, and drip loss at day 3 decreased linearly ($p<0.05$) with the increase in the level of phytoncide

Table 2. Effect of phytoncide on growth performance in finishing pigs

Item	CON ¹	PT2 ¹	PT4 ¹	PT6 ¹	SE ²	p-value		
						Treatment	Linear	Quadratic
BW (kg)								
Initial ³	58.2	58.3	58.2	58.2	0.1	0.80	0.01	0.84
Final ³	116.9 ^b	119.2 ^{ab}	123.0 ^a	114.6 ^b	2.9	0.01	0.70	0.04
0 to 5 week								
ADG (g)	886 ^{ab}	838 ^{ab}	917 ^a	786 ^b	34.4	<0.01	0.63	0.34
ADFI (g)	2,280	2,112	2,135	2,077	65.5	0.43	0.04	0.67
Gain:feed	0.389	0.397	0.430	0.378	0.022	0.76	0.72	0.03
5 to 10 week								
ADG (g)	792	902	934	862	49.3	0.11	0.82	0.01
ADFI (g)	2,829	2,785	2,658	2,520	103.4	0.69	<0.001	0.55
Gain:feed	0.280 ^b	0.324 ^{ab}	0.351 ^a	0.342 ^{ab}	0.014	<0.01	0.24	<0.001
0 to 10 week								
ADG (g)	839 ^{ab}	870 ^{ab}	925 ^a	806 ^b	28.8	0.03	0.47	<0.001
ADFI (g)	2,555 ^a	2,448 ^{ab}	2,397 ^{ab}	2,299 ^b	66.3	0.05	<0.001	0.81
Gain:feed	0.328 ^b	0.355 ^{ab}	0.386 ^a	0.351 ^{ab}	0.015	0.01	0.39	<0.001

SE, standard error; BW, body weight; ADG, average daily gain; ADFI, average daily feed intake.

¹ CON, basal diet; P2, basal diet+0.02% phytoncide; P4, basal diet+0.04% phytoncide; P6, basal diet+0.06% phytoncide.

² Each mean represents 40 observations per treatment.

³ Initial, the 1st day of experiment; final, the last day (d 70) of experiment.

^{ab} Means in the same row with different superscripts differ ($p < 0.05$), initial BW was used as a covariate.

supplementation, and WHC increased linearly ($p < 0.05$) with the increased level of phytoncide supplementation.

DISCUSSION

As an alternative feeding strategy to growth promoters, PFA have received considerable attentions recently. Various trials have demonstrated positive effects of PFA in swine and poultry (Wang et al., 2008; Windisch et al., 2008; Van Krimpen et al., 2010). Wang et al. (2008) reported that 0.04% dietary phytoncide feed additive, which was made of essential oils, flavonoids, pungent substances, and mucliages, did not affect the ADFI of lactating sows. Whereas, Zhang et al. (2012) observed that weaning pigs

fed with 0.2% phytoncide showed a greater G:F ratio. Moreover, Yan et al. (2010) reported that 0.01% essential oil used as PFA increased ADG and G:F at different nutrient densities in pigs. Consistent with these previous studies, our study demonstrated that 0.04% phytoncide increased ADG and G:F ratio in finishing pigs. Essential oils, as a main content of phytoncide additive, may have beneficial effects such as antiviral and antimicrobial activity, and are capable of stimulating enzyme activity and immune function in livestock (Williams and Losa, 2001; Yan et al., 2010). Also, positive of essential oils on digestive system and stimulating effect on the output of digestive enzymes from the pancreas, gut mucosa, and increase bile flow have been reported (Jamroz and Kamel, 2002; Jamroz et al., 2005;

Table 3. Effect of phytoncide on blood profile in finishing pigs

Item	CON ¹	PT2 ¹	PT4 ¹	PT6 ¹	SE ²	p-value				
						Treatment	Time	Treatment×time	Linear	Quadratic
Cortisol (µg/dL)										
Initial ³	1.20	1.20	1.13	1.00	0.24	<0.01	<0.01	0.57	0.08	0.57
Final ³	2.20 ^a	1.00 ^c	1.20 ^{bc}	2.00 ^{ab}	0.27				0.82	0.02
Red blood cell (10 ⁶ /µL)										
Initial ³	6.59	6.43	6.64	6.82	0.34	<0.01	0.36	0.44	0.63	0.46
Final ³	4.67 ^b	6.63 ^{ab}	7.15 ^a	6.54 ^{ab}	0.31				0.71	<0.001
White blood cell (10 ³ /µL)										
Initial ³	22.69	21.68	22.08	21.77	2.47	0.09	0.03	0.64	0.27	0.14
Final ³	19.17	19.47	22.10	17.89	1.42				0.52	0.01

¹ CON, basal diet; PT2, basal diet+0.02% phytoncide; PT4, basal diet+0.04% phytoncide; PT6, basal diet+0.06% phytoncide.

² Standard error. Each mean represents 20 observations per treatment.

³ Initial, the 1st day of experiment; final, the last day (d 70) of experiment.

^{ab} Means in the same row with different superscripts differ ($p < 0.05$).

Table 4. Effect of phytoncide on meat quality in finishing pigs

Item	CON ¹	PT2 ¹	PT4 ¹	PT6 ¹	SE ²	p-value		
						Treatment	Linear	Quadratic
Sensory evaluation								
Color	1.88 ^b	2.17 ^a	1.96 ^{ab}	2.21 ^a	0.077	0.05	0.14	0.57
Marbling	1.79	1.63	1.75	1.92	0.134	0.12	0.40	0.03
Firmness	2.25 ^a	1.96 ^{ab}	1.92 ^b	1.92 ^b	0.10	0.02	0.05	0.11
Lean meat area (cm ²)	44.78	46.84	42.52	44.21	1.97	0.23	0.23	0.42
pH	5.24	5.25	5.32	5.23	0.04	0.71	0.07	0.10
Meat color								
Lightness (L*)	55.98	56.25	57.33	56.45	1.13	0.57	0.14	0.29
Redness (a*)	21.08	20.05	18.38	17.08	1.13	0.12	0.01	0.57
Yellowness (b*)	10.60	9.45	8.83	8.53	0.72	0.26	0.04	0.37
Water holding capacity (WHC, %)	56.94 ^c	64.73 ^b	64.93 ^{ab}	68.18 ^a	1.03	<0.01	0.02	0.76
Drip loss (%)								
3 d	10.66	10.34	9.21	8.65	1.37	0.35	0.03	0.39
6 d	11.82	13.53	11.72	10.26	1.65	0.40	0.17	0.09

¹ CON, basal diet; PT2, basal diet+0.02% phytoncide; PT4, basal diet+0.04% phytoncide; PT6, basal diet+0.06% phytoncide.

² Standard error. Each mean represents 10 observations per treatment.

^{a,b,c} Means in the same row with different superscripts differ (p<0.05).

Jang et al., 2007), which may explain the improvement of growth performance of pigs in our present experiment. Also, specific capabilities of phytochemicals is to enhance the activities of digestive enzymes (bile, pancreatic lipase and amylase etc.) and nutrient absorption (Windisch et al., 2008), which may be another explanation for our results.

The quality of plant materials, selection of particular plants, and forms for administration (i.e., extracts, oils, and dried herbs) also influence their effects on pig performance. As mentioned above, the active substances in phytoncide have a potential capacity to intercalate into the bacterial membrane, disintegrate the membrane, and cause ion leakage, it means that PFA could inhibit the biosynthesis or expression of aflatoxin, or block the enzymatic activity of a biosynthetic enzyme, which including superoxide dismutase, xanthine oxidase, glutathione peroxidase, and two microsomal reductases (Burt, 2004; Holmes et al., 2008; Hashemi and Davoodi, 2010). Those may be the reason why it could promote growth performance of pigs by protecting feed quality. However, inconsistent results have been obtained regarding the effects of PFA on ADFI in animals. Yan et al. (2011a) noted that 1 g/kg herbal extract (*Houttuynia cordata* or *Taraxacum officinale*) used as PFA increased ADFI during a 10-week trial. Wang et al. (2007) and Wenk et al. (2003) reported that herbs used as PFA could improve the flavor and palatability of feed, subsequently increasing the total feed intake and growth performance. Yan et al. (2010) demonstrated that 0.01% essential oil, including thyme, rosemary, oregano extracts, and kaolin covered by starch had a tendency to lower ADFI. One reasonable explanation for the lower ADFI might be the tranquilizing effect of PFA, which reduced the

spontaneous physical activity of the pigs. Physical activity influences heat production, which can cause additional expenditure of energy (Wang et al., 2007). Moreover, pungent substances of some plants (e.g., capsaicin, allitricin, and piperine), which are the active components of PFA, may reduce the palatability of feed and restrict their use for animal feeding purpose (Zhong et al., 2011). In line with our study, previous studies also showed dose-related depressions of palatability of pigs fed with essential oils from the herbs thyme and oregano, as well as from caraway and fennel (Jugl-Chizzola et al., 2006; Schone et al., 2006). Therefore, according to these previous studies the elevate effect of essential may depressed as pungent substances concentration increased.

Czech et al. (2009) reported that pigs fed a mixture with a supplement of 8 g/kg herbal mixture during the whole fattening period were characterized by a significantly higher value of red blood cell system indices, including serotonin, hemoglobin, and RBC. Yan et al. (2011b) confirmed this result, describing increased RBC concentrations in pigs fed 250 or 500 mg/kg of herb extract mixture. Pungent substances (e.g. garlic, black pepper, and chilli), the active components of PFA, could activate blood circulation and metabolic processes (Rodas, 2006). Consistent with that, Iranloye (2002) showed that feeding rats with 200 mg/kg garlic juice daily increased the RBC and WBC. Moreover, Wang et al. (2008) observed that RBC and WBC were increased significantly with PFA treatment. Phytoncide had also been shown to reduce stress responses by lowering stress-related hormones (Kawakami et al., 2004). Collectively, these findings indicate that phytoncide as PFA can increase blood circulation, metabolic processes, and the

immune system of pigs.

Phytoncide can reduce stress responses in stroke-prone spontaneously hypertensive rats (Kawakami et al., 2004). Nam and Uhm (2008) documented that phytoncide decreased serum cortisol levels significantly in college students. Psychological stress affects immune function and predicts infectious disease susceptibility in humans and animals (Marsland et al., 2002). Plant extracts could stimulate T cell-mediated immune responses (Islam et al., 2004). Additionally, flavonoids, one bioactive substance within phytoncide, exhibit inhibitory effects against multiple viruses and improve immunity (Critchfield et al., 1996). In line with previous studies, the concentration of stress-related hormone, cortisol, was reduced significantly with phytoncide supplementation in our present study, indicating that stress was reduced and immunity was enhanced with phytoncide supplementation (Koopmans et al., 2005). In contrast, the effects of PFA on blood profiles have been inconsistent, which might be attributable to interactions between PFA and different feed formulae.

Unacceptable water-holding capacity costs the meat industry millions of dollars annually. Product weight losses due to purge can average as much as 1% to 3% in fresh retail cuts (Offer and Knight, 1988a) and can be as high as 10% in pale soft exudative products (Melody et al., 2004). In addition to the loss of salable weight, purge loss also entails the loss of a significant amount of protein (Offer and Knight, 1988b). On average, purge can contain approximately 112 mg of protein per mL of fluid, mostly water-soluble, sarcoplasmic proteins (Savage et al., 1990). In the current study, phytoncide treatments showed beneficial effects on WHC. It is clear that early postmortem events including the rate and extent of pH decline, proteolysis, and even protein oxidation, are key to influencing the ability of meat to retain moisture (Huff-Lonergan and Lonergan, 2005). Consequently, the antioxidant activities of phytoncide may be a factor causing the higher WHC. Moreover, cortisol was known to increase associated with stress, and stress would decrease meat quality (Terlouw, 2005), in line with our study, cortisol decreased and meat quality increased with 0.04% of phytoncide inclusion.

In conclusion, our results indicate that Korean pine extract supplementation can promote growth performance and elevate meat quality. So that dietary Korean pine extract, phytoncide, could be considered as a potential natural growth promoter in finishing pigs.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

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