Effects of Dietary *Bacillus*-based Probiotic on Growth Performance, Nutrients Digestibility, Blood Characteristics and Fecal Noxious Gas Content in Finishing Pigs

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ABSTRACT: This study was conducted to evaluate the effects of supplementation with *bacillus*-based probiotic (*Bacillus subtilis*, 1.0×10⁷ CFU/g; *Bacillus coagulans*, 2.0×10⁶ CFU/g and *Lactobacillus acidophilus*, 5.0×10⁶ CFU/g) on finishing pigs growth performance, nutrients digestibility, blood characteristics and fecal noxious gas content and to determine the optimal addition level of this probiotic preparation. A total of forty eight pigs with an initial body weight (BW) of 90.60±2.94 kg were allotted to three dietary treatments (four pigs per pen with four pens per treatment) according to a randomized complete block design. Dietary treatment included: 1) CON (basal diet); 2) BP1 (basal diet+ *bacillus*-based probiotic 0.1%) and 3) BP2 (basal diet+ *bacillus*-based probiotic 0.2%). The experiment lasted 6 weeks. Through the entire experimental period, ADG was improved by 11% (p<0.05) in pigs fed diets supplemented with 0.2% *bacillus*-based probiotic compared to pigs fed the basal diet. ADFI and gain/feed were not affected by the treatments (p>0.05). Supplementation of *bacillus*-based probiotic did not affect either DM and N digestibilities or blood characteristics (p>0.05) of pigs. Fecal ammonia nitrogen (NH₃-N) measured at the end of experiment was reduced (p<0.05) when pigs were fed the diet with 0.2% *bacillus*-based probiotic. Fecal acetic acid concentration also decreased significantly (p<0.05) whereas propionic acid concentrations were not affected (p>0.05) when pigs were fed diets with added *bacillus*-based probiotic. In conclusion, dietary supplementation of *bacillus*-based probiotic can increase growth performance and decrease fecal noxious gas content concentration. *(Asian-Aust. J. Anim. Sci. 2006. Vol 19, No. 4 : 587-592)*

Key Words: Probiotics, Digestibility, Blood Characteristics, Fecal Noxious Gas, Finishing Pigs

INTRODUCTION

The intestinal microflora plays a crucial role for both human and animal. During the last few decades, probiotics which also called DFM (Direct-Fed Microbials) have been demonstrated to be useful in maintaining intestinal ecosystem and improving animal health. Recent concerns regarding the use of probiotics suggested its primary action modes were: competition for receptors on the gut mucosa and nutrients with pathogens (competitive exclusion), production of antibacterial substances and stimulation of intestinal immune responses (Vandenbergh, 1993; Dunne et al., 1999; Adlerberth et al., 2000; Wenk, 2000). Although those mechanisms have been well documented by many authors, the efficacy of probiotic preparations in practice is highly inconsistent. Some authors reported the improvement of growth performance (Nousiainen and Setala, 1993), nutrient digestibility (Maxwell et al., 1983), immune system (Fernandes and Shahani, 1990; Malin et al., 1996) and reduction of fecal noxious gas emission (Hong et al., 2002) by the dietary supplementation of probiotics. However, there were also some reverse results obtained in practical feeding trials of growing-finish pigs (Kornegay and Risley, 1996). Various results from previous studies may due to several aspects such as age of animals, strain of bacteria and addition level (Bomba et al., 2002). This indicates that it is necessary to evaluate different probiotics preparations due to its dubious efficacy. In our early studies, results showed that the nutrients digestibility and fecal noxious gas emission were affected by the addition lactic acid bacteria (Chen et al., 2005a, b). We hypothesized that nutrients digestibility and the hindgut microflora can be also influenced by dietary supplementation of probiotics which contain *bacillus* species bacteria, consequently decreased the fecal NH₃-N and VFA concentrations. Therefore, the objectives of current study were to investigate the effects of a *bacillus*-based probiotic preparation on growth performance, nutrients digestibility, blood characteristics and fecal noxious gas content in finishing pigs and determine the optimal addition level of this probiotic preparation.

MATERIALS AND METHODS

Experimental design, animals and diets

A total of 48 [(Landrace×Yorkshire)×Duroc] pigs with an initial BW of 90.60±2.94 kg were used to evaluate the effects of *bacillus*-based probiotic (*Bacillus subtilis*, 1.0×10⁷ CFU/g; *Bacillus coagulans*, 2.0×10⁶ CFU/g and *Lactobacillus acidophilus*, 5.0×10⁶ CFU/g) supplementation...
on growth performance, nutrients digestibility, blood characteristics and fecal noxious gas content in finishing pigs. The experimental period lasted 6 weeks. Pigs were allotted on the basis of initial BW to 3 dietary treatments, each treatment was assigned to 4 replicate pens with 4 pigs allotted on the basis of initial BW to 3 dietary treatments, pigs. The experimental period lasted 6 weeks. Pigs were fed diets containing chromic oxide (Cr$_2$O$_3$) for 7 days prior to the collection period. At the end of experiment, fecal grab samples were collected from 2 pigs in each pen for two consecutive days. Then samples were stored in refrigerator at -20°C until analysis. Before chemical analysis, fecal samples were dried at 70°C for 72 h and finely ground to pass through a 1 mm screen. All the fecal samples, along with feed samples, were analyzed for DM and N according to the AOAC procedures (AOAC, 1995). Chromium was analyzed by UV absorption spectrophotometry (Shimadzu, UV-1201, Japan). N was measured using a Leco NS 2000 Nitrogen Analyzer (LECO Corporation, St. Joseph, MI).

At the initiation of experiment, two pigs were randomly chosen from each pen and bled via jugular venipuncture to obtain blood samples for determining WBC, RBC and lymphocyte. Same pigs were bled again at the end of experiment. Blood samples were collected into K$_2$EDTA vacuum tube (Becton Dickinson Vacutainer Systems, Franklin Lakes, NJ) and analyzed by the automatic blood analyzer (ADVIA 120, Bayer, USA).

Fecal grab samples were also collected for analyzing NH$_3$-N and VFA concentrations at the end of experiment. NH$_3$-N concentration was determined according to the method of Chaney and Marbach (1962). The VFA measured in this experiment included acetic acid, propionic acid and butyric acid. Analysis method of VFA was as follow: 2 g of fecal samples were diluted with 8 ml of distilled water and added two drops of concentrated HCl. Then samples were mixed and centrifuged at 17,400 g for 10 min at 4°C. The supernatant was filtered and pipetted in to 2-ml gas chromatography vials. The VFA concentrations were analyzed by gas chromatography (Hewlett Packard 6890 Plus, USA) according to the method of Otto et al. (2003).

**RESULTS AND DISCUSSION**

**Growth performance**

The results of ADG, ADFI and Gain/feed are shown in Table 2. ADG in BP1 treatment tended to increase without significant difference (p<0.05). When Pigs fed 0.2% probiotic-supplemented diets (BP2), ADG was significant higher compared to CON treatment (p<0.05). ADFI and gain/feed were not affected among the treatments (p>0.05). Current results are in agreement with Jonsson and Conway (1992), who reported dietary addition of *Bacillus* species improved growth performance and health of pigs.

<table>
<thead>
<tr>
<th>Table 1. Formula and chemical composition of diets in finishing pigs (as-fed basis)$^1$</th>
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</thead>
<tbody>
<tr>
<td>Ingredients (%)</td>
</tr>
<tr>
<td>---------------------------------------------------------------</td>
</tr>
<tr>
<td>Corn</td>
</tr>
<tr>
<td>Soybean meal</td>
</tr>
<tr>
<td>Rice bran</td>
</tr>
<tr>
<td>Molasses</td>
</tr>
<tr>
<td>Animal fat</td>
</tr>
<tr>
<td>Dicalcium phosphate</td>
</tr>
<tr>
<td>Calcium carbonate</td>
</tr>
<tr>
<td>L-lysine (78%)</td>
</tr>
<tr>
<td>Salt</td>
</tr>
<tr>
<td>Vitamin premix$^3$</td>
</tr>
<tr>
<td>Mineral premix$^3$</td>
</tr>
<tr>
<td>Choline chloride (60%)</td>
</tr>
<tr>
<td>L-threonine (98%)</td>
</tr>
<tr>
<td>Bacillus-based probiotic</td>
</tr>
<tr>
<td>Chemical composition$^5$</td>
</tr>
<tr>
<td>Digestible energy (kcal/kg)</td>
</tr>
<tr>
<td>Crude protein (%)</td>
</tr>
<tr>
<td>Lysine (%)</td>
</tr>
<tr>
<td>Calcium (%)</td>
</tr>
<tr>
<td>Phosphorus (%)</td>
</tr>
</tbody>
</table>

$^1$ Forty eight pigs with an average initial BW of 90.60±2.94 kg.

$^2$ Abbreviations: CON, control diet; BP1, control diet+0.1% *Bacillus*-based probiotic and BP2, control diet+0.2% *Bacillus*-based probiotic.

$^3$ Provided per kg of complete diet: 4,000 IU of vitamin A; 800 IU of vitamin D$_3$; 17 IU of vitamin E; 2 mg of vitamin K$_3$; 4 mg of vitamin B$_6$; 1 mg of vitamin B$_12$; 16 µg of vitamin B$_12$; 11 mg of pantothenic acid; 20 mg of niacin and 0.02 mg of biotin.

$^4$ Provided per kg of complete diet: 220 mg of Cu; 175 mg of Fe; 191 mg of Zn; 89 mg of Mn; 0.3 mg of I; 0.5 mg of Co and 0.3 mg of Se.

$^5$ Calculated values.
Alexopoulos et al. (2004) observed significant improvement when finishing pigs fed diet included probiotic (Bacillus licheniformis and Bacillus subtilis). Our previous study, conducted by Chen et al. (2005a), also observed an improvement when growing pig fed diets supplemented complex probiotic (Lactobacillus acidophilus, Saccharomyces cerevisiae and Bacillus subtilis). However, Kornegay et al. (1990) reported that there was no effect on growth performance by the supplementation of Lactobacillus acidophilus in finishing pigs.

Unlike the diverse results obtained from growing and finishing pigs experiments, many studies of probiotics conducted in nursery pigs found positive effects when diets added probiotic preparations (Lessard and Brissom, 1987; Park et al., 2001). These results indicated that age of pigs is a considerable fact relate to the probiotics efficacy. Stavric and Kornegay (1995) suggested that probiotics are more effective in animals during microflora development or when microflora stability is impaired. They also reported that the improvement of growth performance will often be marginal during optimal rearing and feeding conditions. Data from our experiment showed that the ADG in control group was 717 g, this value was not as higher as values obtained from some other studies. Therefore, it was reasonable that the response of current probiotic presented more effective in our experiment.

**Nutrients digestibility**

Table 3 shows the effects of bacillus-based probiotic on nutrients digestibility in finishing pigs. Digestibility of DM was not affected by the addition of probiotic (p>0.05). N digestibility was increased slightly in BP2 treatment compared to CON treatment, however, there was no significant difference (p>0.05).

<table>
<thead>
<tr>
<th>Items (%)</th>
<th>CON</th>
<th>BP1</th>
<th>BP2</th>
<th>SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>DM</td>
<td>69.48</td>
<td>70.28</td>
<td>70.63</td>
<td>1.74</td>
</tr>
<tr>
<td>N</td>
<td>63.64</td>
<td>63.69</td>
<td>66.11</td>
<td>2.38</td>
</tr>
</tbody>
</table>

**Blood characteristics**

Effects of bacillus-based probiotic supplementation on blood characteristics in finishing pigs are present in Table 4. Obtained data show that there were no influences on RBC, WBC and lymphocyte when diets added bacillus-based probiotic. The capacity of probiotics to influence the
immune system is one of the more recent developments in this field (Bloksma et al., 1979). Some beneficial effects of probiotics on animal’s immune system have been proposed by many authors. Perdigon et al. (1991) suggested *Lactobacillus casei* have immunoadjuvant activity. Takahashi et al. (1998) and Vitini et al. (2000) reported that *Bifidobacterium longum* and other lactic acid bacteria can increase the total amount of intestinal IgA. However, Kluber et al. (1985) reported no effect of *Streptococcus faecium* on the cell-mediated immune response in weaning pigs. Similar results also observed by Apgar et al. (1993). Current results are in agreement with those reports. Likewise, our previous studies also observed no effects of single or complex probiotic preparations on blood characteristics (WBC, RBC and lymphocyte) in growing and finishing pigs (Chen et al., 2005a, b). These inconsistent responses to probiotics supplementation in growing and finishing pig diets may be due to varying age and health status of the experimental animals used. The effects of probiotics proposed to be more effective during young age and under stress conditions such as weaning or dietary changes for piglets (Jonsson and Conway, 1992). Also, our experimental period only lasted 6 weeks, which might be too short to induce any changes in finishing pigs.

**Fecal noxious gas content**

Fecal NH₃-N was decreased significantly (p<0.05) when pigs fed diets supplemented with 0.2% *Bacillus*-based probiotic compared to pigs fed basal diets (Table 5). Acetic acid and propionic acid were not affected by the addition of *Bacillus*-based probiotic. Butyric acid was decreased significantly when pigs fed diets including *Bacillus*-based probiotic than those of pigs fed basal diets (p<0.05).

Fecal noxious gas emission such as NH₃-N, H₂S and VFA has become on of the major air pollution in modern concentrate pig production (van Breemen et al., 1982; Slanina, 1994). High concentrations of ammonia or H₂S can cause hazard effects to humans and animals (Drummond et al., 1980; Crook et al., 1991; Busse, 1993). Ferket et al. (2002) reviewed the nutritional strategies to reduce negative emissions from nonruminants. They suggested that the ultimately fecal noxious gas emission of animals is related to nutrients utilization and intestinal microflora ecosystem. In our study, NH₃-N was decreased about 20% (685 vs. 568 ppm) by dietary addition of 0.2% probiotic. This is in agreement with Hong et al. (2004), who reported a reduction in NH₃-N when finishing pigs fed diets added complex probiotic (*Phicia anomala* ST, *Galactomyces geotrichum* SR59 and *Thiobacillus* sp.). Ji and Kim (2002) observed the ammonia production of pigs was decreased 21% by the addition of complex probiotics (*Bacillus sp.*, *Aspergillus oryzae*, and *Lactobacillus acidophilus*). However, digestibility of N was not increased in present results. Therefore, the reduction of fecal NH₃-N may not result from the nutrients digestibility but the alternation of intestinal microflora or some other reasons of pigs.

Production of certain VFA is also the result of anaerobic microbial fermentation of soluble carbohydrates (Argenzio and Southworth, 1974; Ferket et al., 2002). Mackie et al. (1998) also suggested that VFA originate partly by anaerobic bacteria in the gastrointestinal tract and feces. Our study observed fecal butyric acid decreased while acetic acid and propionic acid without influence when *bacillus*-based probiotic added to diet. O’Neill and Phillips (1992) suggested the proportion of individual VFA to total VFA concentration associate with odor offensiveness. The manure odor quality mainly contributes from long-chain VFA (Spoelstra, 1980). Thus, the decrease in the proportion of long-chain or branched-chanined VFA such as butyrate, isobutyrate and valerate has the potential to reduce odor emission (Ferket et al., 2002).

**IMPLICATIONS**

This study indicates that dietary supplementation of *bacillus*-based probiotic preparation at the level of 0.2% is effective in improving growth performance and reducing fecal NH₃-N and butyric acid concentrations in finishing pigs. Therefore, we propose that current probiotic preparation can replace some double-side effect additives such antibiotics or growth promoters etc. Possible mechanisms need to be further investigated in order to finely explain how this *bacillus*-based probiotic exert its beneficial effects.

**REFERENCES**


Table 5. Effects of *Bacillus*-based probiotic on fecal NH₃-N and VFA concentrations in finishing pigs¹

<table>
<thead>
<tr>
<th>Items</th>
<th>CON²</th>
<th>BP1²</th>
<th>BP2²</th>
<th>SE³</th>
</tr>
</thead>
<tbody>
<tr>
<td>NH₃-N (ppm)</td>
<td>685</td>
<td>650</td>
<td>568</td>
<td>28</td>
</tr>
<tr>
<td>Volatile fatty acids</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acetic acid</td>
<td>373</td>
<td>332</td>
<td>357</td>
<td>38</td>
</tr>
<tr>
<td>Propionic acid</td>
<td>458</td>
<td>360</td>
<td>451</td>
<td>35</td>
</tr>
<tr>
<td>Butyric acid</td>
<td>238</td>
<td>175</td>
<td>180</td>
<td>14</td>
</tr>
</tbody>
</table>

¹ Forty eight pigs with an average initial BW of 9.60±2.94 kg.
² Abbreviations: CON, control diet; BP1, control diet+0.1% *Bacillus*-based probiotic and BP2, control diet+0.2% *Bacillus*-based probiotic.
³ Pooled standard error.

a, b means in the same row with different superscripts differ (p<0.05).


