

Oviduct-specific Glycoprotein 1 Locus is Associated with Litter Size and Weight of Ovaries in Pigs

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ABSTRACT : Oviduct-specific glycoprotein 1 (*OVGP1*) is implicated in playing a role in fertilization and early embryo development. In this study, we have obtained the sequence of intron 9 of *OVGP1* gene in swine. Comparative sequencing of Meishan (a native Chinese breed) and Large White pig breeds revealed an A/T substitution at position 943. A PCR-*EcoRI*-RFLP assay was developed to detect this mutation. Polymorphism analysis in Qingping animals showed that pigs with BB genotype had lower number of piglets born alive (NBA) in multiple parities than pigs with AA ($p < 0.05$) and AB genotype ($p < 0.01$). In Large White×Meishan (LW×M) F₂ offspring, the weight of both ovaries (OW) of the BB genotype was significantly lighter than that of AB ($p = 0.05$) and AA ($p < 0.01$) genotypes. Analysis of the data also revealed that the mutation locus affected these two traits mostly by additive effects. These studies indicated that the polymorphism was associated with NBA and OW in two distinct populations and further investigations in more purebreds or crossbreds are needed to confirm these results. (*Asian-Aust. J. Anim. Sci.* 2006. Vol 19, No. 5 : 632-637)

Key Words : Oviduct-specific Glycoprotein 1, Litter Size, Female Reproductive Tract Components, Pigs

INTRODUCTION

Mammalian oviduct designs a suitable microenvironment for fertilization, early embryonic development and delivery of a viable embryo to the uterus (Boatman et al., 1997). *OVGP1* is a high molecular weight glycoprotein belonging to the chitinase protein family and presented in the oviduct (Agarwal et al., 2002). It is synthesized from the oviductal epithelial cells and secreted into the oviductal lumen during estrogen dominance in the human (Arias et al., 1994; Bhatt et al., 2004). In human and baboons, this glycoprotein is specifically expressed within the oviduct during the perovulatory phase of the menstrual cycle and several investigators indicated that synthesis of the *OVGP1* was controlled by ovarian steroids (Rapisarda et al., 1993; O'Day-Bowman et al., 1996). *OVGP1* can attach itself to the ovulated oocyte and early embryo during their transit in the oviduct (Malette et al., 1995; O'Day-Bowman et al., 1996). Several reports showed that *OVGP1* adhered to the zona pellucida (ZP) of the ovulated egg thereby enhancing the binding of sperm to the ZP and the speed of sperm penetration within the oviduct (O'Day-Bowman et al., 1996; Boatman et al., 1997), while the blocking of *OVGP1* with antibodies significantly decrease species-specific sperm binding onto oocytes *in vitro* (Schmidt et al., 1997). *OVGP1*

in the perivitelline space or endocytosed by the pre-implantation embryo may regulate differentiation during the morula to blastocyst transition (Boatman et al., 1997). All these researches suggested that *OVGP1* might have a role in the early reproductive events occurring within the oviduct. Due to the physiological function of *OVGP1* during fertilization, *OVGP1* is a functional candidate gene for reproductive traits in mammal.

Variations in *OVGP1* gene could be used as genetic markers to select animals for breeding. However, it has not been previously reported how a genetic variant of this gene affects litter size in pigs. The purpose of our research was to determine whether the single nucleotide polymorphisms (SNP) of the ninth intron in porcine *OVGP1* gene would be associated with female reproductive traits.

MATERIALS AND METHODS

Experimental animals

In this study, 44 Meishan pigs (a native Chinese breed), 36 Large white pigs, 66 Line DIV pigs, 127 Large white×Meishan resource family F₂ offspring were bred and raised at the genetic nucleus station owned by Huazhong Agricultural University. Twenty-one Tongcheng pigs and 30 Erhualian pigs were derived from Tongcheng (Tongcheng, Hubei Province) and Houqiao (Xishan, Jiangshu Province) research farms in China, respectively.

The purebred Qingping animals used in this study were born and raised in Qingping research farm of Hubei province in China. At farrowing, total number born (TNB) and the number of piglets born alive (NBA) were recorded.

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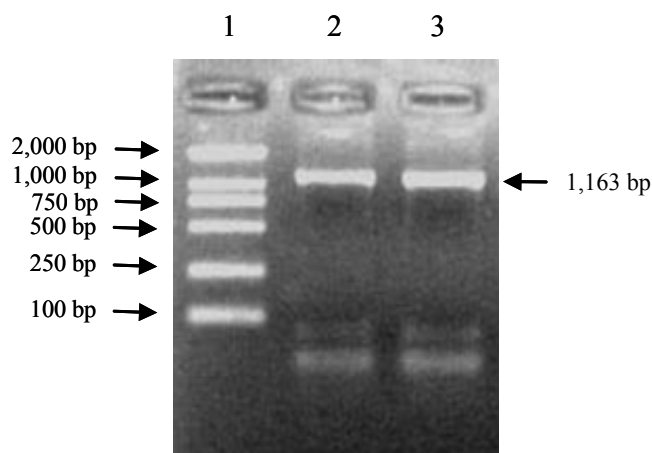


Figure 1. PCR amplified products of intron 9 of *OVGP1* gene. Lane 1: DL 2,000 markers; Lane 2, 3: PCR products in Meishan pigs.

The pig resource family was established by mating three Large White boars to seven Meishan sows. Five males and twenty-three females in the F_1 generation were selected for intercrossing randomly. All the F_2 offspring were slaughtered for testing, including 127 gilts with reproductive tracts components records. Reproductive tract characteristics, including length of uterine horns (LU), uterine weight (UW), length of oviduct (LO) and weight of both ovaries (OW) were recorded. LU was measured according to the method of Lin (1992).

DNA preparation

Genomic DNA was isolated from white blood cells as described by Xiong et al. (1999). After isolation, DNA pellet was dissolved in TE buffer and was stored at -20°C .

Cloning and sequencing of the amplified fragment of intron 9

Primer pair 1 (Forward primer: 5'-AGTGGTTCCTATCTGCCT-3'. Reverse primer: 5'-ACATCATCCAGGTC CAAAGT-3') was developed to amplify a region for the porcine *OVGP1* gene predicted to encompass parts of exons 9, 10 and intron 9 by comparison with the human *OVGP1* cDNA (GenBank accession number: NM 002557) and swine *OVGP1* mRNA (GenBank accession number: NM 214070). PCR was performed in 25 μl reactions containing: 1 \times PCR buffer, 1.5 mM MgCl_2 , 250 $\mu\text{mol/L}$ dNTP, 5 pm of each PCR primer, 2 U *Taq* DNA polymerase (Biostar International, Toronto, Canada), and 200 ng genomic DNA as template. PCR was run in the GeneAmp PCR system 9600 Thermocycler (Perkin Elmer, Foster City, CA, USA) as follows: 94°C for 4 min followed by 35 cycles of 94°C for 1 min, 63°C for 50 s, 72°C for 1 min, and final extension of 10 min at 72°C . The purified PCR products

were cloned into the pMD18-T vector and were sequenced by Sangon Company (Shanghai, China). The sequencing results of different pig breeds were compared by using CLUSTALX software.

Detection of PCR-*EcoRI*-RFLP

Primers 2 were designed based on sequencing result to amplify a 326 bp fragment from intron 9. Forward primer: 5'-AGTGGTTCCTATCTGCCT-3'. Reverse primer: 5'-ACATCATCCAGGTC CAAAGT-3'. PCR amplification (20 μl final volume) was performed using 250 ng genomic DNA, 200 $\mu\text{mol/L}$ dNTP, 5 pm of each primer, and 1 U *Taq* DNA polymerase with the reaction buffer supplied by the manufacturer. The conditions for PCR are as follows: 94°C for 4 min; 33 cycles of 94°C for 30 min, 63°C for 30 s, and 72°C for 1 min. For the PCR-RFLP assays, 8.5 μl of PCR products were digested with 5 units *EcoRI* (TaKaRa, Tokyo, Japan) in 1 \times digestion buffer added in a total volume of 10 μl , following digestion for 4 h at 37°C . Digested products were separated by a 8% polyacrylamide gel electrophoresis (PAGE) in 1 \times TBE buffer and stained with silver. Allelic forms of porcine *OVGP1* intron 9 were identified as B (digested by *EcoRI* into 176 bp and 150 bp fragments) and A (a 326 bp fragment undigested by *EcoRI*).

Statistical analysis

The association between genotype and reproduction traits was performed with the least square method (GLM procedure, SAS version 8.0). The model used to analyze the data from Qingping pigs included effects of the *OVGP1* SNP genotypes and line as fixed effects. The Qingping pig was included in the model as a random. NBA in multiple parities of Qingping individuals were averaged from 2-7 parities. The model used to analyze the data from F_2 gilts included effects of the *OVGP1* SNP genotypes and season of slaughter as fixed effects and F_2 individuals as a random effect.

Both additive and dominance effects were also estimated using REG procedure of SAS version 8.0, where the additive effect was denoted as -1, 0 and 1 for AA, AB and BB, respectively, and the dominance effect represented as 1, -1 and 1 for AA, AB and BB, respectively (Liu, 1998).

RESULTS

Amplification, cloning and sequencing analysis of intron 9

The primer pair 1 produced fragments with genomic DNA from Meishan and Large White pigs. Products of PCR were detected by electrophoresis on 1% agarose gel (Figure 1). Sequence analysis of PCR products revealed that a new *EcoRI* polymorphism was identified at 943th position where the nucleotide A was substituted by nucleotide T.

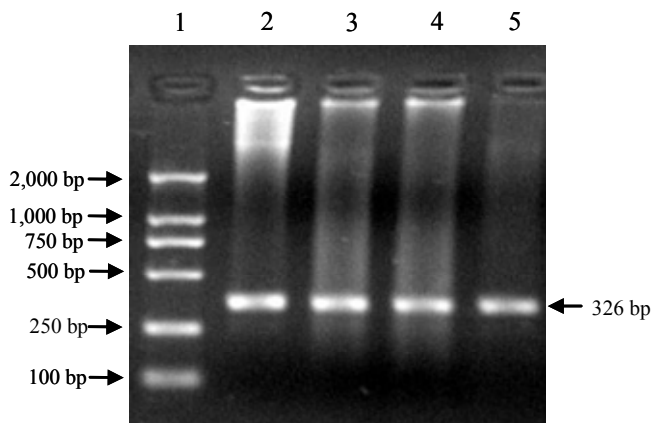


Figure 2. PCR amplified products of intron 9 with primer pair 2. Lane 1: DL 2,000 marker; Lane 2-5: PCR products.

Table 1. Abbreviations of names for traits

No.	Abbreviations	Names	Unit
1	TNB	Total number born	piglet
2	NBA	Number born alive	piglet
3	UW	Uterine weight	g
4	OW	Weight of both ovaries	g
5	LU	Length of uterus horn	cm
6	LO	Length of oviduct	cm

Polymorphism in intron 9

A 326 bp fragment was amplified with primer pair 2. PCR was performed on DNA of seven pig breeds and 124 F₂ offspring. Results of amplification were shown in Figure 2.

The A/T substitution can be detected by PCR-*EcoRI*-RFLP. The PCR products were digested with restriction enzyme (*EcoRI*) and the fragments were separated by electrophoresing on an 8% PAGE. Three genotypes were presented: AA (326 bp), AB (326 bp+176 bp+150 bp), BB (176 bp+150 bp) (Figure 3).

Frequencies of allele and genotype of different pig breeds

Allele frequencies for the PCR-*EcoRI*-RFLP were studied in seven different populations (Table 2). Allele A was the predominant allele in all pig populations and allele B was found only in Chinese indigenous pig breeds, such as

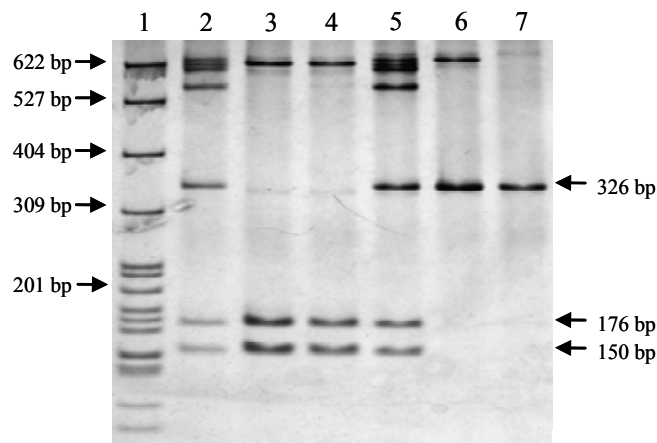


Figure 3. The PCR-RFLP results of intron 9 with primer pair 2. Lane 1: pBR322 DNA/*MspI* marker; Lane 2, 5: genotype AB; Lane 3, 4: genotype BB; Lane 6, 7: genotype AA.

Meishan, Qingping, Tongcheng, Erhualian pigs. There were three genotypes (AA, BB and AB) existed in all Chinese native pig populations except Erhualian animals. Chi square (χ^2) testing showed highly significant difference in genotype frequencies between Large White pig population and Chinese native pig breeds (Meishan breed and Tongcheng breed) (Table 3), indicating clearly genetic differentiation at the locus between Chinese and Western pigs, whereas the differences among Chinese Erhualian, Meishan and Tongcheng pigs were significant as well.

Association analysis

Results of tests for *OVGP1* genotypes and litter size traits on Qingping pigs were given in Table 4. Pigs with *BB* genotype had less NBA than pigs with *AA* genotype ($p < 0.05$) and *AB* genotype ($p < 0.01$), with additive effect -0.897 piglet. Significant association with NBA of multiple parities in Qingping animals was found, but no significant conclusion could be made on other litter size traits. The results also indicated that *AB* sows showed the highest and *BB* sows showed the lowest litter size values in the later and subsequent parities. Table 5 summarized the association of porcine *OVGP1* genotypes with the female reproductive trait components on LW×M pigs. The results showed that the OW of *BB* gilts were significantly lighter than *AB* gilts

Table 2. Distribution of *EcoRI*-RFLP genotype and allele frequencies among different pig populations

Populations	N	Genotype and frequency			Allele frequency	
		AA	AB	BB	A	B
Meishan	68	0.37(25)	0.49(33)	0.15(10)	0.61	0.39
Qingping	92	0.61(56)	0.32(29)	0.08(7)	0.77	0.23
Tongcheng	21	0.38(8)	0.52(11)	0.10(2)	0.64	0.36
Erhualian	30	0.97(29)	0.03(1)	0.00(0)	0.98	0.02
Line DIV	66	1.00(66)	0.00(0)	0.00(0)	1.00	0.00
Large White	36	1.00(36)	0.00(0)	0.00(0)	1.00	0.00
Duroc	15	1.00(15)	0.00(0)	0.00(0)	1.00	0.00

N: Total number of pigs observed.

Table 3. χ^2 test results for the genotype frequency distribution among different populations

Populations	Qingping	Tongcheng	Erhualian	Line DIV	Large White	Duroc
Meishan	9.260	0.375	30.224 ^{**}	61.456 ^{**}	38.812 ^{**}	19.682
Qingping		3.746	13.754	33.447 ^{**}	19.599	8.846
Tongcheng			21.328 [*]	48.034 ^{**}	28.870 ^{**}	14.534
Erhualian				2.223	1.218	0.511
Line DIV					0	0
Large White						0

* $p < 0.05$; A, B, ** $p < 0.01$.

Table 4. Effect of OVGP1 EcoRI-RFLP genotypes on litter size traits in Qingping sows

Trait	Least square means \pm standardized error			Additive effect	Dominant effect	
	AA	AB	BB			
N	56	29	7			
Multiple parities	TNB	11.003 \pm 0.264	11.450 \pm 0.369	10.143 \pm 0.812	-0.328 \pm 0.400	-0.396 \pm 0.268
	NBA	10.775 \pm 0.238 ^a	11.024 \pm 0.330 ^A	8.982 \pm 0.667 ^{BB}	-0.897 \pm 0.358*	-0.573 \pm 0.240*

N: Total number of pigs observed.

Letter denoting significant difference between groups: a, b, * $p < 0.05$; A, B, ** $p < 0.01$.

Table 5. Effect of OVGP1 EcoRI-RFLP genotypes on reproductive tracts components traits in LW \times M F₂ offspring

Traits	Least square means \pm standardized error			Additive effect	Dominant effect
	AA	AB	BB		
N	39	67	18		
UW (g)	429.615 \pm 41.985	400.270 \pm 35.195	454.167 \pm 61.800	14.038 \pm 29.351	28.163 \pm 19.500
LU (cm)	108.987 \pm 8.022	98.821 \pm 6.167	100.833 \pm 11.809	4.077 \pm 6.894	4.525 \pm 4.580
OW (g)	15.719 \pm 1.078 ^A	13.437 \pm 0.840 ^{AB}	12.156 \pm 1.481 ^B	1.723 \pm 0.904	0.463 \pm 0.622
LO (cm)	20.473 \pm 0.834	19.144 \pm 0.625	21.059 \pm 1.230	-0.218 \pm 0.722	0.658 \pm 0.475

N: Total number of pigs observed.

Letter denoting significant difference between groups: a, b, * $p < 0.05$; A, B, ** $p < 0.01$.

($p = 0.05$) and AA gilts ($p < 0.01$), the additive effect was 1.723 g ($p = 0.05$). Significant association with OW in F₂ gilts was observed as well. No significant association was found for other reproductive tract characteristics.

DISCUSSION

In this study, the candidate gene approach has been used to demonstrate associations between specific genes and litter size. Using this approach, polymorphisms in some defined genes such as the estrogen receptor (Legault et al., 1996; Rothschild et al., 1996; Short et al., 1997; Li et al., 2004), follicle-stimulating hormone- β (Zhao et al., 1998; Li et al., 2002), prolactin receptor (Vincent et al., 1998) and retinol-binding protein (Rothschild et al., 2000) have all been reported to be associated with litter size in swine. So candidate gene approach is a very efficacious method to find molecular markers.

OVGP1 gene has been extensively investigated in humans. In human the highest expression of human OVGP1 at the time of ovulation is consistent with a supportive role in fertilization and early embryo development (Lok et al., 2002). In pigs, it has been reported that OVGP1 significantly reduced the incidence of polyspermy among pig eggs matured and fertilized *in vitro* (Andrew et al.,

2000). In addition, OVGP1 provided a significant increase in postcleavage development from embryo to blastocyst in swine (Andrew et al., 2000). Therefore, this gene may play an important role in the *in vivo* fertilization process (Rapisarda et al., 1993; Malette et al., 1995; Andrew et al., 2000). Thus, in our study OVGP1 gene was selected as a candidate gene to be associated with reproductive trait in swine.

The genetic effect on reproductive traits was investigated in Qingping sows and F₂ offspring animals. Based on the data representing litter records from Qingping sows, the NBA and TNB in multiple parities were higher for AB sows than for the sows of the other two genotypes, though on association between the OVGP1 gene locus and TNB was discovered. In view of these, it is tempting to suggest that AB sows have better performance than BB sows for litter size traits in Qingping pigs.

However, our results in F₂ offspring showed that piglets with AA genotypes had heavier OW than AB. Association between the marker and the traits may vary across populations, lines or families. This was shown in several studies with the ESR gene for reproductive traits. According to Rothschild et al. (1996) and Short et al. (1997) the B allele was advantageous to the A allele for litter size. Another research showed no statistically significant effect

of the ESR genotype on litter size (Legault et al., 1996). The observed difference between Qingping sows and F₂ gilts may be explained that there are variations in the genetic background. In addition, the observed effects might be caused by the linkage of this locus with other quantitative trait locus (QTLs) which contributed to the reproductive traits.

Mutation of nucleotide can alter gene function, either by changing the coding region of the protein, translation or stability of the mRNA or control of transcription of the gene. In the experiments reported here, a polymorphism analyzed in *OVGP1* gene was found in intron 9. Although the SNP in introns do not directly alter any amino acid residue, they may play a role in regulating gene expression and thus their constituent SNPs may be directly related to functional variation (Zhang et al., 2005). Furthermore, it should be taken into consideration that the location of *OVGP1* gene on chromosome has not been reported in the literature, and hence we cannot say whether *OVGP1* is the gene leading to different litter size and OW itself or if it is only a genetic marker linked with other QTLs which contributed to the reproductive traits.

This is the initial step to consider *OVGP1* gene as a candidate gene for litter size. This polymorphism could be a potential genetic marker for reproductive traits, especially for NBA. The presented estimations are based on the data from Qingping breeds and F₂ intercross pedigrees, however, in this study the populations are weak and the number of Qingping individuals or F₂ gilts is limited. So, analyzing more animals is necessary to confirm the association between the *OVGP1* genotype and reproductive traits in Qingping pigs or other purebreds and crossbreds.

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